

respectively. The older hatchling appeared healthy, whereas the second hatchling had suffered damage to the right eye from bites inflicted by red imported fire ants (*Solenopsis invicta*). Landers et al. (1980) also documented attacks by fire ants on hatchling gopher tortoises and theorized that they may be a significant mortality factor.

Because pet and free-ranging dogs were common in the vicinity of the nest, personnel of Audobon Zoological Garden in New Orleans agreed to head-start the hatchlings for 1.5 - 2 years. The young tortoises will be released into suitable habitat on a protected site in Louisiana.

In addition to the resident female, the owners of this site reported seeing another large tortoise in and near the burrow excavated by the female during late spring or early summer 1988. However, I thoroughly searched the area within 300 m of the female's burrow and was not able to locate any additional adult-sized burrows.

The second case of recent tortoise reproduction in Louisiana was documented from Bush, St. Tammany Parish. The owner of this site noted two young tortoises and three small burrows 23 September; two more small burrows were located the following day. On 14 October, I visited the site and confirmed her identifications. One active adult-sized burrow and several inactive burrows were located on the site. The dimensions of the active burrow, presumed to be that of the female parent, were 30 x 16 cm. During my visit, I observed five active hatchling burrows (mean entrance dimensions 6.3 x 2.5 cm. and one inactive start; young were present in at least four of the five active burrows. Three of the burrows had been excavated at the base of some large object, such as a wood pile; the other two burrows were in the open. Total burrow lengths were about 25 cm (two burrows), 30 cm, and 45 cm (two burrows). All burrows were excavated at a shallow angle (ca. 15 degrees); thus, the burrow terminus was generally only 2-4 cm below the soil surface. Distances from the adult burrow to the hatchling burrows were 5.0, 7.8, 17.5, 22.9, and 70.1 m. I was unable to locate egg shell fragments or any other sign of the nest site. The resident female was uniquely marked, and the owners of the site reported occasional observations of non-resident adult tortoises on their property. However, I was unable to locate any active adult-sized burrows within 300 m of the presumed female's burrow.

These observations indicate that the gopher tortoise is not functionally

extinct in Louisiana. Discovery of reproduction by apparently isolated female tortoises raises questions about gopher tortoise biology in need of investigation. Studies are needed to determine home range sizes of male tortoises under low-density conditions, and whether gopher tortoise populations can persist in non-"colonial" situations.

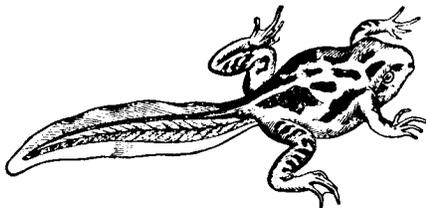
I would like to acknowledge Gary Lester, Louisiana Natural Heritage Program, for initiating the recent gopher tortoise surveys in Louisiana.

LITERATURE CITED

- Auffenberg, W. and R. Franz. 1982. The status and distribution of the gopher tortoise *Gopherus polyphemus*. R. B. Bury (ed.), North American tortoises: conservation and ecology. U. S. Fish and Wildl. Ser. Res. Rep. 12.
- Douglas, J. F. 1978. Refugia of juvenile gopher tortoises, *Gopherus polyphemus* (Reptilia, Testudines, Testudinidae). J. Herpetol. 12: 413 - 415.
- Jennings, R. D. and T. H. Fritts. 1983. The status of the gopher tortoise, *Gopherus polyphemus* Daudin. Final report submitted to U.S. Fish and Wildlife Service, Jackson, Mississippi. 15 pp.
- Landers, J. L., J. A. Garner, and W. A. McRae. 1980. Reproduction of gopher tortoises (*Gopher polyphemus*) in southwestern Georgia. Herpetologica 36:353-361.
- Lohofener, R. and L. Lohmeier. 1984. The status of *Gopherus polyphemus* (Testudines, Testudinidae) west of the Tombigbee and Mobile Rivers. A report presented to U.S. Fish and Wildlife Service along with a petition to list the western population of the gopher tortoise. 126 pp.

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THE SAVANNAH RIVER ECOLOGY LABORATORY HERPETOLOGICAL MUSEUM

Herpetological research is a major emphasis at the University of Georgia's Savannah River Ecology Laboratory (SREL) in South Carolina (Gibbons 1977; Gibbons and Caldwell 1980; Gibbons and Patterson 1978). In recognition of the useful information that preserved reptiles and amphibians can provide for systematic, ecological and evolutionary studies, SREL maintains a representative museum collection of these animals. The scope of the collection is largely regional, but a variety of specimens from other states and countries are available. The purpose of the collection is three-fold. First, maintenance of a synoptic collection of reptiles and amphibians found on the Savannah River Plant (SRP). Second, to provide verification of important locality records with voucher specimens, and third, to provide interested researchers with a comprehensive regional collection for systematic, ecological and evolutionary study. The objective of this note is to familiarize the herpetological community with the amphibian and reptile collection at SREL.

Specimens entered into the collection date from 1950. However, specimens were not collected on the SRP and catalogued until 1968. Collection and preservation of specimens was in accordance with guidelines given in ASIH et al. (1987). Presently, the collection holds over 3000 catalogued alcoholic specimens representing 30 families, 80 genera, and 165 species. Of these, approximately 2000 are amphibians. The representation of major orders is shown in Figure 1. Major strengths of the collection include large series of local amphibians and small colubrid snakes, and an extensive collection of larval amphibians. Traffic volume on the 200 km of paved roads on the SRP is light, and road-killed specimens are rarely hit more than once. As a result, large numbers of freshly killed snakes have been collected and incorporated into the museum (Kaufman and Gibbons 1975). Regionally notable specimens include a significant range extension for *Rhadinaea flavilata* (Young 1988), and a collection of *Kinosternon baurii* from the western limit of their distribution in South Carolina (Lamb 1983a, b). Catalogued entries encompass 17 states with over 2600 records for South Carolina.

The entire collection was inventoried in 1988 and a computer file was created. Researchers interested in lists

of our holdings for various taxonomic groups or geographic regions can obtain them by writing to the Acting Curator of Herpetology. Loan policies are liberal, but visits are encouraged.

In short, we hope that research-oriented herpetologists will take advantage of the excellent regional collection maintained at SREL.

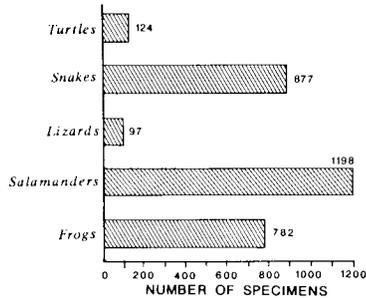


Figure 1. Number of catalogued entries for major orders of amphibians and reptiles in the Savannah River Ecology Laboratory Museum. Totals do not include lot catalogued entries.

LITERATURE CITED

- ASIH, HL, and SSAR. 1987. Guidelines for use of live amphibians and reptiles in field research. Jointly published by Amer. Soc. Ichthyol. Herpetol., Herpetologists' League, and Soc. Stud. Amphib. Rept. 14 pp.
- Gibbons, J. W. 1977. Snakes of the Savannah River Plant with information about snakebite prevention and treatment. Environmental Research Park Publication. 26 pp. SRO-NERP-1.
- Gibbons, J. W. and J. Caldwell. 1980. Herpetology at the Savannah River Ecology Laboratory. Herp. Rev. 11:72-74.
- Gibbons, J. W. and K. K. Patterson. 1978. The reptiles and amphibians of the Savannah River Plant. Environmental Research Park Publication. 24 pp. SRO-NERP-2.
- Kaufman, G. A. and J. W. Gibbons. 1975. Weight-length relationships in thirteen species of snakes in the southeastern United States. Herpetologica 3:31-37.
- Lamb, T. 1983a. On the problematic identification of *Kinosternon* (Testudines: Kinosternidae) in Georgia, with new state localities for *Kinosternon bauri*. Georgia J. Sci. 41:115-120.
- Lamb, T. 1983b. The striped mud turtle (*Kinosternon bauri*) in South Carolina, a confirmation through multivariate character analysis. Herpetologica 39:383-390.

Young, D. P. 1988. Geographic distribution: *Rhadinaea flavilata*. SSAR Herp. Review 19:20.

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FLORIDA STATE MUSEUM CHANGES ITS NAME BUT KEEPS ITS ACRONYM

The Florida State Museum was founded in 1917 by an act of the state legislature as a department of the University of Florida. FSM became the official acronym for citing specimens in print and remained in effect until 1946, when the museum temporarily closed its doors upon the retirement of its long time director, Dr. T.H. van Hyning. In 1951 the museum reopened and almost immediately, several collections maintained by the Department of Biology were transferred to its care, greatly increasing the museum's holdings. As a result of these transfers, the acronym was changed to UF, which remains in effect to this day.

A sign outside the museum reads "J.C. Dickinson, Jr. Hall, Florida State Museum" (Dr. Dickinson was the third director). However, another sign on the building declares "State Museum of Florida." Dr. Peter Bennett, fifth and present director, noted this discrepancy and also began meeting people in his travels who thought the Florida State Museum was located in Tallahassee and was part of Florida State University. He resolved to stop this confusion and, in June, 1988, the governor signed a name change into law. The Florida State Museum will now be called the Florida Museum of Natural History.

The logical new replacement acronym for UF would be FMNH, but it is already in use by the Field Museum of Natural History. In any event, abandoning UF would only add to the diversity of incorrect acronyms already present in the literature. For example, authors still commonly use FSM. UF/FSM and, less frequently, UF-FSU are also seen. Leviton et al. (1985) list both UF and UF-FSU as official acronyms for the Florida State Museum. Unfortunately, they don't mention in either their Parts 1 or 2 that UF-FSU refers only to a part of the Ichthyology Division collection, which was received from Florida State University (the Ichthyology Division is presently computerizing its collection and has dropped the UF-FSU acronym).

The Florida Museum of Natural History is one of the few institutions for which its acronym does not match its name. Although the present acronym has been in effect for 38 years, the new museum name will undoubtedly result in new incorrect acronyms in the literature. Authors should read the institutional loan form, on which the correct acronym is written (until recently, UF/FSM was on the Herpetology Division form!) or, in gen-

eral, consult the appropriate person at the institution before submitting a manuscript.

LITERATURE CITED

Leviton, A.E., R.H. Gibbs, Jr., E. Heal, and C.E. Dawson. 1985. Standards in herpetology and ichthyology: Part I. Standard symbolic codes for institutional resource collections in herpetology and ichthyology. Copeia 1985:802-832.

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UNUSUAL NEST SITE OF A SCINCID LIZARD *Sphenomorphus* *kinabalensis* FROM SABAH, MALAYSIA

Sphenomorphus kinabalensis is an endemic Bornean skink, known only from high altitude areas around Mt. Kinabalu, Sabah. Since Bartlett (1895) described this species, no information concerning its natural history has been published. In the present paper, we report an egg deposition site observed during a zoological survey in Sabah. Voucher specimens were deposited in the herpetological collection of the Department of Zoology, Kyoto University.

Sixteen eggs, covered by white parchment-like shells, were found in a nest of a ponerine ant, *Anochetus princeps*. The ant nest was made in tunnels (probably made by passalid beetles) in a fallen tree on the floor of the moss-forest at Ranau (near Mt. Kinabalu, alt. ca. 1300 m), Sabah, on 13 August 1987. All eggs were half-buried in the wet substrate of a brood chamber, where numerous larvae and pupae of the ant also were observed. We collected 14 eggs (elliptical, 9.8 X 7.9 - 13.8 X 9.4 mm), and kept them on wet paper in a plastic cup at an air temperature of 22°-32° C. Six of the eggs produced hatchlings, 20.1+27.5 - 21.5+28.3 mm snout to vent (SVL)+tail lengths, whereas the remainder molded. Exact dates of hatching were: 18 (N=1), 19(1), 24(1), 29(1), and 30(2) August 1987. We also examined five other fallen trees with similar tunnels in the same forest. However, neither lizard eggs nor ant nests were found.

We dissected seven adult female *S. kinabalensis* (SVL: 45.0-57.4 mm) collected near Mt. Kinabalu in August 1979, and examined their gonads. Of the seven animals, six possessed two oviducal eggs or yolked follicles, and the other a single oviducal egg.

The number of eggs found in the ant nest were probably the contribution of eight or more clutches, and thus, it is likely that the skinks specifically selected the ant nest as a communal oviposition site. The lack of eggs in other trees with similar tunnels but without ant nests support this assumption.